

Periodic Acid Schiff (PAS) Diastase Stain Kit

Description: The Periodic Acid Schiff (PAS) Diastase Stain Kit is intended for use in histological demonstration of lymphocytes and mucopolysaccharides. The α -Amylase digestion step acts on glycogen to break it into smaller sugars that are then washed off the tissue section allowing visual comparison of digested and undigested slides. The PAS reaction in tissue sections is useful for the demonstration of mucopolysaccharides.

PAS Positive Material: Magenta
Nuclei: Blue

Uses/Limitations: Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.


Control Tissue: Liver

Ordering information regarding individual components on back page!

Kit Contents:


<u>Item #</u>	<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
AAS030	Alpha-Amylase Solution (1%)	30 ml	2-8° C
PAQ030	Periodic Acid Solution	30 ml	2-8° C
SRF030	Schiff's Solution	30 ml	2-8° C
HMM030	Hematoxylin, Mayer's	30 ml	18-25° C
BRT030	Bluing Reagent	30ml	18-25° C

Precautions: Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Storage: 2° C  25° C


**Mixed Storage Conditions.
Separate Contents.**

Doc: IFU-TemplateMixedStorageev2



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Instructions For Use

PAD-2-IFU

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Revision: 1

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Procedure:


1. Deparaffinize two identical sections if necessary and hydrate to distilled water.
 2. If sections are Zenker-fixed, remove mercuric chloride crystals using iodine and clear with sodium thiosulfate. Rinse in running tap water.
 3. Apply 5-10 drops of Alpha-Amylase Solution (1%) to one slide and incubate for 10-30 minutes at room temperature.
 4. Rinse in 2 changes of distilled water.
- Note: The remainder of this procedure is performed on both the “digested” and “undigested” slides.
5. Apply 5-10 drops of Periodic Acid Solution (1%) to tissue section and incubate for 5 minutes.
 6. Rinse slide in 4 changes of distilled water.
 7. Apply 5-10 drops of Schiff's Solution to tissue section and incubate for 10-20 minutes.
 8. Rinse slide in warm running tap water for 2 minutes.
 9. Rinse slide in distilled water.
 10. Apply 5-10 drops of Hematoxylin, Mayer's (Lillie's Modification) to tissue section and incubate for 1 minute.
 11. Rinse in running tap water for 1 minute followed by 2 changes of distilled water.
 12. Apply 5-10 drops of Bluing Reagent for 5 seconds and rinse in distilled water.
 13. Dehydrate through graded alcohols.
 14. Clear, and mount in synthetic resin.

References:

1. Culling CFA, Allison RT, Barr WT.: Cellular Pathology Technique, 4th Edition. Butterworths, Pages 216-220, 1985.
2. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. CV Mosby, Columbus, OH. Pages 164-167, 1980.


Bulk Reagent Ordering Information and Current Pricing at www.scytek.com


Description:	Catalog #	Volume
Alpha-Amylase Solution (1%)	AAS250	250 ml
	AAS500	500 ml
	AAS999	1000 ml
Periodic Acid Solution	PAQ250	250 ml
	PAQ500	500 ml
	PAQ999	1000 ml
Schiff's Solution	SRF250	250 ml
	SRF500	500 ml
	SRF999	1000 ml
Hematoxylin, Mayer's (Lillie's)	HMM500	500 ml
	HMM999	1000 ml
	HMM3800	1 Gallon
Bluing Reagent	BRT500	500 ml
	BRT999	1000 ml
	BRT3800	1 Gallon

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