

# P.T.A.H. Stain Kit for Microwave

## (Phosphotungstic Acid Hematoxylin)

### Description:

The P.T.A.H. Stain Kit for Microwave is intended for use in the histological visualization of collagen, striated muscle, glial fibers and collagen without using Zenker's Fixative with Mercuric Chloride as a mordant. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Fibrin, Striated Muscle, Glial Fibers:	Blue
Collagen:	Brownish/Red
Nuclei:	Blue

### Uses/Limitations:

Not to be taken internally.  
 For In-Vitro Diagnostic use only.  
 Histological applications.  
 Do not use past expiration date.  
 Use caution when handling reagents.  
 Non-Sterile.

### Control Tissue:

Striated Muscle

### Availability/Contents:

<u>Item #</u>	<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
ZCS030	Zinc Chloride Solution (10%)	30 ml	18-25°C.
FAS030	Ferric Ammonium Sulfate Sol.	30 ml	18-25°C.
HPA030	P.T.A.H. Solution	30 ml x 2	18-25°C.

### Storage:

Store at 18-25°C.

### Precautions:


Avoid contact with skin and eyes.  
 Harmful if swallowed.  
 Follow all Federal, State, and local regulations regarding disposal.

### Equipment Needed: 500 Watt Microwave Oven

### Procedure (Microwave):

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Place slide to fresh distilled water for 1 minute.
3. Fill a coplin jar approximately 80% full with DI water.
4. Place coplin jar in microwave and heat until hot but not boiling.

Storage: 18° C  25° C



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5. Carefully place slide across the top of the coplin jar and apply 5 drops of Zinc Chloride Solution (10%) and heat in microwave for 10 seconds. Leave jar with slide in the microwave during the incubation period to better maintain temperature.

6. Incubate slide for 15 minutes.

**NOTE:** Water in coplin jar will maintain reagent temperature during staining procedure.

7. Rinse slide in running tap water for 1 minute.

8. Rinse in distilled water for 1 minute.

**Note:** During rinse step, reheat water in coplin jar to hot but not boiling.

9. Carefully place slide across the top of the coplin jar and apply 5 drops of Ferric Ammonium Sulfate Solution and heat in microwave for 10 seconds. Leave jar with slide in the microwave during the incubation period to better maintain temperature.

10. Incubate slide for 2 minutes.

11. Rinse slide in running tap water for 2 minutes.

12. Rinse in distilled water for 1 minute.

**Note:** During rinse step, reheat water in coplin jar to hot but not boiling.

13. Carefully place slide across the top of the coplin jar and apply 5 drops of Phosphotungstic Acid Hematoxylin Solution and heat in microwave for 10 seconds. Leave jar with slide in the microwave during the incubation period to better maintain temperature.

14. Incubate slide for 15 minutes.

15. Shake off Phosphotungstic Acid Hematoxylin Solution and repeat Steps 13 and 14.


16. Differentiate section in 95% Reagent Alcohol. Check section using microscope for proper differentiation.


17. Dehydrate in 3 changes of Absolute Alcohol.

18. Clear in 3 changes of fresh Xylene or Xylene Substitute, and mount in synthetic resin.

## References:


- Shapiro, S.H., Sohn, L.C.; Rapid Microwave Phosphotungstic Acid-Hematoxylin Stain for Paraffin and Glycol Methacrylate Sections; The Journal of Histotechnology; Volume 17, Number 2, June 1994, pages 125-126.

Storage: 18° C  25° C



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