

Instructions For Use A00004-IFU-IVD

Rev. Date: June 29, 2021

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Bcl-2; Clone 124

Catalog Number	Format	Volume
A00004-0002	(Ready-To-Use)	2 ml
A00004-0007	(Ready-To-Use)	7 ml
A00004-0025	(Ready-To-Use)	25 ml
A00004-C.1	(Concentrate)	0.1 ml
A00004-C	(Concentrate)	1 ml

Intended Use

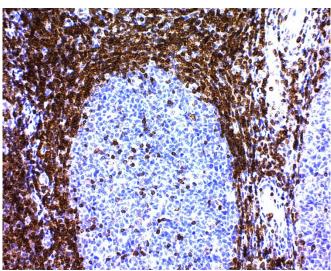
For In Vitro Diagnostic use. This antibody is intended for the qualitative visualization of the anatomical elements listed in the Specificity section. It is intended to be used within an Immunohistochemistry (IHC) procedure on formalin-fixed paraffin-embedded (FFPE) human tissue followed by visualization by light microscopy. Any diagnostic interpretation of the results of this antibody is to be complemented by morphological studies using proper controls and should be evaluated within the context of the patient's clinical history and other diagnostic tests by a qualified pathologist.

Description

Titer/Working Dilution	Ready-to-Use: No further dilution required.
y	Concentrate: Suggested dilution is 1:100-300
Species:	Mouse
Immunogen:	BALB/C mice were immunized with synthetic peptide sequence
Ū	comprising amino acids 51-54 of bcl-2 protein.
Clone:	124
Isotype:	lgG1, Kappa.
Entrez Gene ID:	596 (Human)
Hu Chromosome Loc.:	
Synonyms:	Apoptosis regulator Bcl-2, B-cell CLL/lymphoma-2
Mol. Wt. of Antigen:	25-26kDa
Format:	Ready-To-Use antibody has been pretitered and quality controlled to work on formalin-fixed paraffin-embedded as well as acetone fixed cryostat tissue sections. No further titration is required.
	Concentrate antibody is provided at 200µg/ml of Ab purified from Bioreactor Concentrate by Protein A/G. Prepared in 10mM PBS with 0.05% BSA & 0.05% Sodium Azide.
Specificity:	This antibody reacts with a 25 kD bcl protein, which lies within the cell rather than on the cell surface. It stains neoplastic cells of follicular lymphoma, hairy cell leukemia, high grade B and T cell lymphomas, lymphoblastic lymphomas and anaplastic large cell lymphoma.
Background:	Expression of Bcl-2 alpha oncoprotein inhibits programmed cell death (apoptosis). In most follicular lymphomas, neoplastic germinal centers express high levels of Bcl-2 alpha protein, whereas the normal or hyperplastic germinal centers are negative. Consequently, this antibody is valuable when distinguishing between reactive and neoplastic follicular proliferation in lymph node biopsies. It may also be used in distinguishing between those follicular lymphomas that express Bcl-2 protein and the small number in which the neoplastic cells are Bcl-2 negative.
Species Reactivity: Positive Control:	Human, Others-not known Tonsil or follicular lymphomas. Jurkat, K562, HL-60, or HeLa
	Cells.
Cellular Localization:	Outer mitochondrial membranes and endoplasmic reticulum as well as nuclear membranes.
Microbiological State:	



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Human tonsil stained using Bcl-2; Clone 124. Results were visualized using ScyTek's UHP500 detection system and DAB Chromogen/Substrate Kit (High Contrast) Cat# ACV500.

Materials and Reagents Required but not Provided

- 1. Control tissue and reagents
- 2. Xylene, graded alcohols, and deionized/distilled water
- 3. Antibody Diluent.

4. IHC detection system. Suggested: ScyTek Cat# ABZ125 "CRF Anti-Polyvalent HRP Polymer" and ScyTek Cat# ACV500 "DAB Chromogen/Substrate Kit (High Contrast)".

- 5. Wash buffer for rinses (ScyTek Cat# TBT500)
- 6. HIER Retrieval Solution
- 7. Hematoxylin counterstain and bluing reagent (ScyTek Cat# HMM500 and BRT500)
- 8. Mounting medium and coverslips

Note: ScyTek Laboratories has a wide range of IHC reagents and ancillaries that can be found at scytek.com.

Procedure

1. Tissue Section Pretreatment (Required): Staining of formalin fixed, paraffin embedded tissue sections is significantly enhanced by pretreatment with pH 8-9 HIER Solution (see ScyTek catalog# ETA or TES for instructions).

2. **Primary Antibody Incubation Time:** We suggest an incubation period of 30 minutes at room temperature. However, depending upon the fixation conditions and the staining system employed, optimal incubation should be determined by the user.

3. Visualization: For maximum staining intensity we recommend the "CRF Anti-Polyvalent HRP Polymer" (ScyTek catalog# ABZ125, see IFU for instructions) combined with the "DAB Chromogen/Substrate Bulk Pack (High Contrast)" (ScyTek catalog# ACV500, see IFU for instructions).

Storage and Stability

Do not Freeze. Store at 2-8°C. Return to 2-8° immediately after use. Do not use after expiration date printed on label. Verify visually that antibody has not been contaminated before use. Do not use if reagent becomes cloudy or precipitates.



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Limitations

Immunohistochemistry is a complex technique involving both histological and immunological detection methods. Tissue processing and handling prior to immunostaining can cause inconsistent results. Variations in fixation and embedding or the inherent nature of the tissue specimen may cause variations in results. Endogenous peroxidase activity or pseudoperoxidase activity in erythrocytes and endogenous biotin may cause non-specific staining depending on detection system used. This data sheet's recommendations and procedures were validated using ScyTek IHC reagents and may not be suitable for other detection systems.

Precautions

1. Contains Sodium Azide as a preservative (0.09% w/v), do not ingest. Sodium Azide may react with lead and copper plumbing to form highly explosive metal azides. Upon disposal, flush with large volumes of water to prevent azide build-up in plumbing. This product contains no hazardous material at a <u>reportable concentration</u> according to U.S. 29 CFR 1910.1200, OSHA Hazardous Communication Standard and EC Directive 91/155/EC. 2. Do not pipette by mouth.

3. Avoid contact of reagents and specimens with skin and mucous membranes.

Avoid microbial contamination of reagents or increased nonspecific staining may occur.
The user must validate any procedures and recommendations that differ from this data sheet.

6. The SDS may be found at scytek.com

References

1. Cleary et al. Cell 47: 19, 1986.

2. Tsujimoto et al. Proc Natl Acad Sci (USA) 83: 5214, 1986.

3. Hockenbery et al. Nature 348: 334, 1990.

4. Pezzella et al. Am J Pathol 137: 225, 1990.

5. Tsuchido K, Yamada M, Satou T, Otsuki Y, Shimizu SI, Kobayashi H. Cytology of sclerosing epithelioid fibrosarcoma in pleural effusion. Diagnostic cytopathology. 2010 Oct;38(10):748-53.

 Gurlek U, Abakay CD, Ozkan L, Saraydaroglu O, Kurt M, Cetintas SK. The evaluation of bcl-2 expression as a prognostic marker in early stage laryngeal cancer. Tumori Journal. 2013 Nov;99(6):682-8.

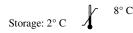
7. Büyüktaş D, Örnek S, Tokat F, Tecimer T, Ferhanoğlu B. IRF4-Rearranged Large B-Cell Lymphoma on Waldeyer's Ring: A Case Report. Turkish Journal of Hematology. 2020 Dec;37(4):292.

Warranty

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