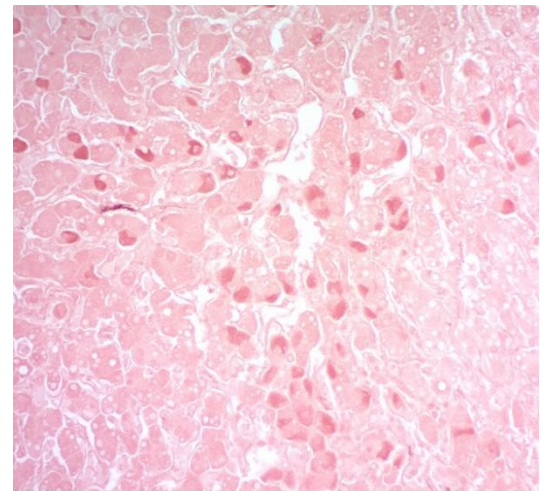


Orcein Stain Kit

(For Hepatitis B and Elastic Fibers)

Description: The Orcein Stain may be used in histology procedures for the visualization of Hepatitis B surface Antigen (HBsAg), elastic fibers, and copper associated proteins. HBsAg appears as irregular shaped aggregates in the cytoplasmic region of the cells. This reagent may be used on formalin-fixed, paraffin-embedded sections.

HBsAg:	Dark Red/Brown
Elastic Fibers:	Dark Red/Brown
Copper Assoc. Proteins:	Dark Red/Brown
Background:	Light Reddish/Purple



Hepatitis B Surface Antigen in Human Liver at 20X stained with the Orcein Stain Kit.

Uses/Limitations: Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.

Control Tissue: Known hepatitis positive liver.
Lung for elastic fiber.
Any well fixed tissue cut 3-5 microns.


Ordering information regarding additional recommended individual components on back page!

Availability/Contents:


<u>Item #</u>	<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
PPG030	Potassium Permanganate Sol. (5%)	30 ml	18-25°C
SAQ030	Sulfuric Acid Solution (3%)	30 ml	18-25°C
OQB030	Oxalic Acid Solution (2%)	30 ml	18-25°C
OAA030	Orcein Solution	2 x 30 ml	18-25°C
DSA030	Differentiating Solution	30 ml	18-25°C

Precautions: Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Storage: 18° C  25° C

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Procedure:**Prepare Oxidizing Immediately Prior to Beginning Procedure:**

Combine: 50 ml Distilled Water
 5 ml Potassium Permanganate Solution (5%)
 3 ml Sulfuric Acid Solution (3%)

Mix thoroughly.

1. Deparaffinize sections if necessary and hydrate to distilled water.
2. Incubate slide in freshly prepared Oxidizing Solution for 10 minutes.
3. Rinse slide briefly in running tap water followed by 1 dip in distilled water.
4. Apply 3-5 drops of Oxalic Acid Solution (2%) to tissue section and incubate slide for 10 minutes.

Note: Section should be colorless following this step.

5. Rinse slide for 1 minute in running tap water followed by 2 dips in distilled water.
6. Incubate slide in coplin jar containing Orcein solution for 4-8 hours (2 hours is sufficient for elastin). **Note:** Ensure tissue is fully immersed in staining jar. Close lid to prevent evaporation.
7. Rinse slide in Alcohol, Reagent (70%).
8. Differentiate in Differentiating solution for 10-60 seconds.
9. Dip slide in Alcohol, Reagent (70%) and check slide microscopically for proper differentiation.

Note: Repeat step 8 if necessary.

10. Dehydrate quickly in 3 changes of absolute alcohol.
11. Clear, and mount in synthetic resin.


Note: If darker staining is preferred:

- 1) Incubation time in Orcein solution may be increased.
and/or
- 2) Differentiation may be omitted by replacing steps 7-9 with a simple rinse with deionized water.

References:

1. Deodhar K.P., Tapp E., Scheuer P.J. Orcein staining of Hepatitis B Antigen in paraffin sections of Liver Biopsies. Journal of Clinical Pathology; vol. 28: pages 66-70, 1975.
2. Salaspuro, M., Sipponen, P. Demonstration of an intracellular copper-binding protein by Orcein staining in long-standing cholestatic liver diseases. Gut, 1976, volume 17: pages 787-790.

Storage: 18° C  25° C



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Instructions For Use HBK-2-IFU

Rev. Date: Dec. 7, 2018


Revision: 2


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
Bulk Reagent Ordering Information and Current Pricing at www.scytek.com

Description:	Catalog #	Volume
Potassium Permanganate Solution (5%)	PPG030	30 ml
	PPG125	125 ml
Sulfuric Acid Solution (3%)	SAQ125	125 ml
	SAQ500	500 ml
	SAQ999	1000 ml
Oxalic Acid Solution (2%)	OQB500	500 ml
	OQB999	1000 ml
Orcein Solution	OAA125	125 ml
	OAA500	500 ml
	OAA999	1000 ml
Differentiating Solution	DSA125	125 ml
	DSA500	500 ml
Alcohol, Reagent (70%)	EAS500	500 ml
	EAS999	1000 ml

Storage: 18° C  25° C

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