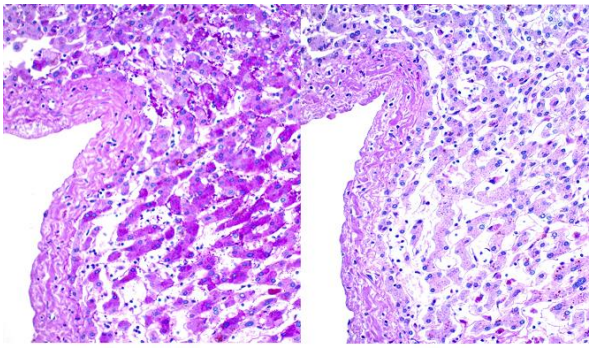
P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 – Tel. (435) 755-9848 - Fax (435) 755-0015 - www.scytek.com

Alpha Amylase Solution (1%)

Description: This α-Amylase reagent acts on glycogen to break it into smaller sugars that are then washed off the tissue section allowing visual comparison of digested and undigested slides. This reagent is usually used within a PAS staining procedure.

Results (PAS staining):

PAS Positive Material:	Magenta
Nuclei:	Blue
Glycogen:	Removed



Undigested Glycogen Digested

Uses/Limitations:
Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.

Control Tissue: Liver

Suggested PAS Kit Contents (ScyTek Cat# PAD-1):

Item #	Kit Contents	Volume	Storage
AAS250	Alpha-Amylase Solution (1%)	250 ml	2-8° C
PAQ250	Periodic Acid Solution	250 ml	2-8° C
SRF250	Schiff's Solution	250 ml	2-8° C
HMM125	Hematoxylin, Mayer's	2x125 ml	18-25°C
BRT125	Bluing Reagent	2x125ml	18-25°C

Precautions:
Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Storage: Store reagent at 2-8°C.

Storage: 2° C 8° C

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Suggested PAS Procedure (ScyTek Reagents):

1. Deparaffinize two serial sections if necessary and hydrate to distilled water.
2. If sections are Zenker-fixed, remove mercuric chloride crystals using iodine and clear with sodium thiosulfate. Rinse in running tap water.
3. Apply Alpha-Amylase Solution (1%) to one slide and incubate for 10-30 minutes at room temperature.
4. Rinse in 2 changes of distilled water.

Note: The remainder of this procedure is performed on both the “digested” and “undigested” slides.

5. Apply Periodic Acid Solution (1%) to tissue section and incubate for 5 minutes.
6. Rinse slide in 4 changes of distilled water.
7. Apply Schiff’s Solution to tissue section and incubate for 10-20 minutes.
8. Rinse slide in warm running tap water for 2 minutes.
9. Rinse slide in distilled water.
10. Apply Hematoxylin, Mayer’s (Lillie’s Modification) to tissue section and incubate for 1 minute.
11. Rinse in running tap water for 1 minute followed by 2 changes of distilled water.
12. Apply Bluing Reagent for 5 seconds and rinse in distilled water.
13. Dehydrate through graded alcohols.
14. Clear, and mount in synthetic resin.

References:

1. Culling CFA, Allison RT, Barr WT.: Cellular Pathology Technique, 4th Edition. Butterworths, Pages 216-220, 1985.
2. Sheenan, D.C., Hrapchak, B.B. Theory and Practice of Histotechnology, 2nd Edition. CV Mosby, Columbus, OH. Pages 164-167, 1980.