

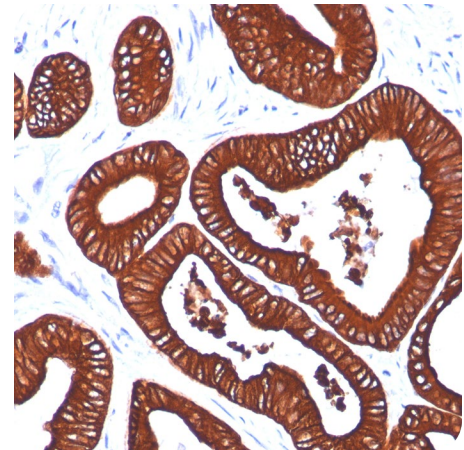
DAB Substrate Buffer

Description:

The DAB Substrate buffer has been developed for applications that require excellent contrast between the chromogen and Hematoxylin counterstain. When in the presence of peroxidase enzyme and DAB chromogen, the substrate helps to produce a brown precipitate that is insoluble in alcohol. The use of liquid components reduces some risks associated with handling powders (ie. dust inhalation), and eliminates waste which often results from using tablets that require a predetermined final volume. This product provides slightly less sensitivity than ScyTek's DAB Substrate (High Sensitivity) catalog#: ACU.

Uses/Limitations:

Not to be taken internally.
For In Vitro Diagnostic use.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.



Control Tissue:

Any well fixed paraffin embedded or frozen tissue section.

Ordering information regarding individual components on back page!

Availability:

<u>Item #</u>	<u>Component</u>	<u>Volume</u>	<u>Storage</u>
ACC500	DAB Substrate	500 ml	2-8°C
ACC999	DAB Substrate	1000 ml	2-8°C


Bulk Reagent Ordering Information and Current Pricing at www.scytek.com


Precautions:

Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Required but not included:

DAB Chromogen (Scytek Catalog# ACB)

Storage: 2° C  8° C

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Procedure:

1. Rinse slide in deionized water prior to application of DAB mixture.
2. Combine 50-80µl of DAB Chromogen with 1ml of DAB Substrate and apply to tissue for 5 minutes.



NOTE: Range = 50-80µl of DAB Chromogen / 1 ml of DAB Substrate. 50µl / ml produces optimal staining quality, 80µl/ml produces maximum sensitivity. Combined mixture may be used for up to six hours.

OPTIONAL: For additional sensitivity, add 20µl of DAB Enhancer (cat.# ACM030, not included) to each 1ml of mixed solution.

3. Rinse slide using deionized water.
4. Apply mixture a second time and incubate for another 5 minutes.
5. Rinse slide in two changes of deionized water.
6. Counterstain as desired.
7. Dehydrate through graded alcohol, clear and coverslip in synthetic medium.

References:

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3. Oznurulu, Y., Celik, I., Sur, E., Telatar, T., Ozparlak, H. Comparative Skin Histology of the White New Zealand and Angora Rabbits: Histometrical and Immunohistochemical Evaluations. *Journal of Animal and Veterinary Advances*. 2009, Volume 8(9): pages 1694-1701.
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5. Gupta, P., Dhingra, K.K., Singhal, S., Khurana, N., Mandal, S., Mohta, A. Primary primitive neuroectodermal tumour (PNET) of testis: an unsuspected diagnosis. *Pathology*, February 2010, Volume 42, Issue 2: pages 179-181. Doi: 10.3109/00313020903494029.
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7. Ozaltin, F., Ibsirlioglu, T., et al. Disruption of PTPRO Causes Childhood-Onset Nephrotic Syndrome. *The American Journal of Human Genetics*. July 15, 2011, Volume 89(1): pages 139-147. Doi: 10.1016/j.ajhg.2011.05.026.

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