

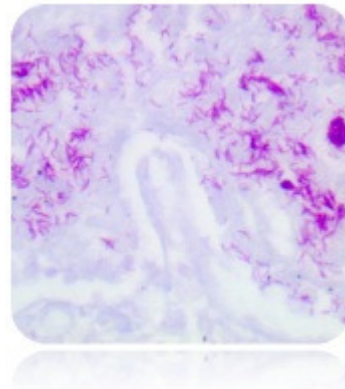
Fite's Stain Kit

(For Leprosy and Nocardia)

Description: The Fite's Stain Kit (For Leprosy and Nocardia) is intended for use in the histological visualization of mycobacterium Lepra bacillus and Nocardia. This kit may be used on formalin-fixed, paraffin-embedded or frozen sections.

Lepra bacillus:	Red
Nocardia:	Red
Background:	Blue

Uses/Limitations: Not to be taken internally.
For In-Vitro Diagnostic use only.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.




Control Tissue: TCS0009-5 (5 Slides)
TCS0009-25 (25 Slides)
Any well fixed paraffin embedded
Nocardia or Lepra bacillus infected
tissue.


Ordering information regarding individual components on back page!

Kit Contents:


<u>Item #</u>	<u>Kit Contents</u>	<u>Volume</u>	<u>Storage</u>
XPO125	Xylene-Peanut Oil Solution	125 ml	18-25 °C
CFZ125	Carbol Fuchsin Solution	125 ml	18-25 °C
AAM500	Acid Alcohol Solution (1%)	500 ml	18-25 °C
MBS125	Methylene Blue Solution	125 ml	18-25 °C

Precautions: Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Storage: 18° C  25° C

 ScyTek Laboratories, Inc.
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Lepra bacillus Procedure (Standard):




1. Deparaffinize sections in 2 changes of Xylene-Peanut Oil Solution for 12 minutes each.
2. Air dry slide for 15 minutes “without” removing oil film covering tissue section. Remaining film prevents de-staining of Lepra bacillus during differentiation.
3. Rinse slide in several changes of distilled water.
4. Incubate slide in Carbol Fuchsin Solution for 15 minutes.
5. Rinse slide in several changes of distilled water.
6. Differentiate section in Acid Alcohol Solution (1%) until background is pink.
7. Rinse slide in distilled water and check by microscope for correct differentiation.
8. Rinse in running tap water for 1 minute followed by 1 rinse in distilled water.
9. Dip slide 2-3 times in Methylene Blue Solution.
10. Dip slide quickly in distilled water and check by microscope for correct staining.
11. Air dry slide at room temperature.
12. Dip slide several times in Xylene or Xylene Substitute.
13. Mount in synthetic resin.

Nocardia Procedure:**Preparation of Reagents Prior to Beginning:**

1. Prepare **Diluted Acid Alcohol Solution** by mixing 25ml of Acid Alcohol Solution (1%) with 25ml of Distilled Water.

Procedure:

1. Deparaffinize sections in 2 changes of Xylene-Peanut Oil Solution for 12 minutes each.
2. Air dry slide for 15 minutes “without” removing oil film covering tissue section. Remaining film prevents de-staining of Lepra bacillus during differentiation.
3. Rinse slide in several changes of distilled water.
4. Incubate slide in Carbol Fuchsin Solution for 15 minutes.
5. Rinse slide in several changes of distilled water.
6. Dip slide 2-3 times in Diluted Acid Alcohol Solution.
7. Rinse slide in distilled water and check by microscope for correct differentiation. Avoid decolorizing the Nocardia organism.
8. Rinse in running tap water for 1 minute followed by 1 rinse in distilled water.
9. Dip slide 2-3 times in Methylene Blue Solution.
10. Dip slide quickly in distilled water and check by microscope for correct staining.
11. Air dry slide at room temperature.
12. Dip slide several times in Xylene or Xylene Substitute.
13. Mount in synthetic resin.


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References:

1. Echeverri, C., et al. Fite Stain Positivity in Rhodococcus equi: Yet Another Acid-Fast Organism in Respiratory Cytology – A Case Report. Diagnostic Cytopathology; April 2001, Volume 24, Issue 4, pages 244-246.
2. Crowder, C., Taylor, HW., Modified Fite Stain for Demonstration of Mycobacterium Species in Tissue Sections; Journal of Histotechnology; 1996, Volume 19; 2: pages 133-134.
3. Mallory, Pathological Technique; page 275.

Bulk Reagent Ordering Information and Current Pricing at www.scytek.com


Description:	Catalog #	Volume
Xylene-Peanut Oil Solution	XPO125	125 ml
	XPO500	500 ml
	XPO999	1000 ml
Carbol Fuchsin Solution	CFZ125	125 ml
	CFZ500	500 ml
	CFZ999	1000 ml
Acid Alcohol Solution (1%)	AAM500	500 ml
	AAM999	1000 ml
Methylene Blue Solution	MBS125	125 ml
	MBS500	500 ml
	MBS999	1000 ml

 Storage: 18° C  25° C



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