

# Instructions For Use JSK-IFU

205 South 600 West Logan, Utah 84323, U.S.A. – Tel. (800) 729-8350 – Tel. (435) 755-9848 – Fax (435) 755-0015 – www.scytek.com Rev. 2, 8/2/2022

# Jones Methenamine Silver Stain Kit

(For Basement Membrane)

# **Description and Principle**

The Jones Stain Kit is intended for use in histological demonstration of the basement membrane and reticular fibers. This procedure is ideal for staining renal glomerular basement membranes. The main function of the basement membrane and reticular fibers is to provide anchorage and support. They are normally found throughout the body, particularly in the kidney, spleen, and lung.

Basement membranes and reticular fibers are demonstrated by methenamine silver through oxidation of carbohydrates to aldehydes. The aldehydes formed by oxidation bind silver ions from the methenamine silver complex and reduce the silver to its metallic form.

#### **Expected Results**

Basement Membrane: Black
Oxidizable Carbohydrates: Black
Nuclei: Red
Cytoplasm: Light Pink

Kit Contents	Storage
1. Periodic Acid Solution	2-8°C
2. Methenamine Solution	2-8°C
3. Silver Nitrate Solution (5%)	2-8°C
4. Borax Solution	18-25°C
5. Gold Chloride Solution (0.2%)	2-8°C
6. Sodium Thiosulfate Solution (5%)	18-25°C
7. Nuclear Fast Red Solution (Enhanced)	18-25°C

## Suggested Controls (not provided)

Kidney cut at 2 microns, Lung, Spleen

# **Uses/Limitations**

For In-Vitro Diagnostic use only.

Do not use if reagents become cloudy or precipitate

Do not use past expiration date.

Use caution when handling reagents.

Non-Sterile

Intended for FFPE sections cut at 5-10  $\mu m. \,$ 

This procedure has not been optimized for frozen sections.

Frozen sections may require protocol modification.

# <u>Storage</u>

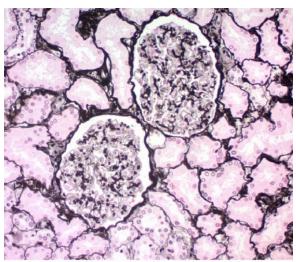
Store kit contents according to individual bottle labels

### Safety and Precautions

Please see current Safety Data Sheets (SDS) for this product and components GHS classification, pictograms, and full hazard/precautionary statements.

#### **Important Notes**

- $\overline{1}$ . All glassware used in this procedure should be chemically cleaned and rinsed thoroughly in distilled water.
- 2. Do <u>not</u> use metal forceps to remove slides from reagents. Use plastic forceps only.
- 3. Equilibrate all reagents to room temperature prior to use.
- 4. Increasing the number of slides present in working Methenamine Silver Solution will decrease the rate of silver staining. Monitor slides during Methenamine Silver incubation to achieve desired intensity.



Normal Kidney glomerular basement membrane cut at 2 micron thickness. Stained for 1 hour with working Silver Methenamine Solution. Toned for 30 seconds with Gold Chloride Solution (0.2%). Magnification 20X.

#### **Procedure**

- 1. Deparaffinize sections and hydrate to distilled water.
- 2. Incubate slide in Periodic Acid Solution for 15 minutes.
- 3. Rinse in 2 changes of distilled water.
- 4. Combine the following for a working Methenamine Silver Solution:

42 ml Methenamine Solution 2.5 ml Silver Nitrate Solution (5%) 6 ml Borax Solution

#### Note: Mixed solution may not be stored for reuse.

5. Place working Methenamine Silver Solution in 65° centigrade water bath and allow temperature to equilibrate.

6. Incubate slide in working Methenamine Silver Solution for 20-60 minutes. Dip slide in "hot" distilled water and check under a microscope for evaluation of silver impregnation. Basement membrane and reticular fibers should be black. If color is not sufficient, return the slide to working Methenamine Silver Solution checking frequently until desired staining intensity is achieved.

- 7. Rinse in 4 changes of distilled water.
- 8. Tone slide in Gold Chloride Solution (0.2%) for 30 seconds.
- 9. Rinse in 4 changes of distilled water.
- 10. Incubate slide in Sodium Thiosulfate Solution (5%) for 2 minutes.
- 11. Rinse in tap water followed by 2 changes of distilled water.
- 12 Incubate slide in Nuclear Fast Red Solution for 2 minutes.
- 13. Rinse slide quickly in distilled water.
- 14. Rinse slide using absolute alcohol.
- 15. Dehydrate in 3 changes of absolute alcohol, clear, and mount in synthetic resin.

# References

- 1. Carson, F.L., (1997), Histotechnology; A Self instructional text, second edition. ASCP Press, Chicago, IL. Pages 151-154
- 2. Jones, D. B. (1957). Nephrotic glomerulonephritis. The American journal of
- Jones, D. B. (1957). Nephrotic glomerulonephritis. The American journal of pathology, 33(2), 313.
   Kunak, C. S., Ugan, R. A., Cadirci, E., Karakus, E., Polat, B., Un, H., ... & Karaman, A. (2016). Nephroprotective potential of carnitine against glycerol and contrast-induced kidney injury in rats through modulation of oxidative stress, proinflammatory cytokines, and apoptosis. The British Journal of Radiology, 89(1058), 20140724.
   Saritemur, M., Un, H., Cadirci, E., Karakus, E., Akpinar, E., Halici, Z., ... & Atmaca, H. T. (2015). Tnf-α inhibition by infliximab as a new target for the prevention of glycerol-contrast-induced nephropathy. Environmental toxicology and pharmacology, 39(2), 577-588.
   Ugan, R. A., Cadirci, E., Halici, Z., Toktay, E., & Cinar, I. (2018). The role of urotensin-II and its receptors in sepsis-induced lung injury under diabetic
- urotensin-II and its receptors in sepsis-induced lung injury under diabetic conditions. European journal of pharmacology, 818, 457-469.

ScyTek Laboratories, Inc. 205 South 600 West Logan, UT 84321 435-755-9848 U.S.A.



EC REP

Emergo Europe Prinsessegracht 20 2514 AP The Hague, The Netherlands