P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 – Tel. (435) 755-9848 - Fax (435) 755-0015 - www.scytek.com

SensiTek Alk-Phos Anti-Polyvalent Lab Pack

Description:

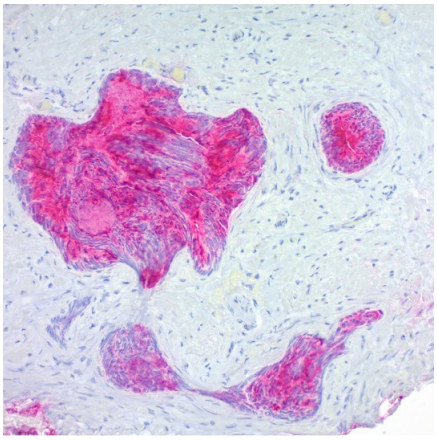
The SensiTek staining kit provides unmatched sensitivity with incubation times of 20 minutes each for the Link Antibody and Enzyme Label. The bulk kits are ideal for high volume laboratories. Each Pack contains one bottle of Super Block (universal protein block), one bottle of Biotinylated Antibody (Polyvalent), and one bottle of Alkaline Phosphatase Labeled Streptavidin. These Lab-Packs provide an extremely economical alternative for automated staining systems and we encourage you to evaluate the addition of this in your current system.

Species of Origin: Goat
Antigen Specificity: Anti-Polyvalent (Mouse, Rat, Rabbit and Guinea Pig).
Preadsorbed Against: Human
Enzyme Conjugate: Alkaline Phosphatase
Chromogen Substrate: None Provided

Contains:
One container of Super Block.
One container of Anti-polyvalent.
One container of Alkaline Phosphatase.

Uses/Limitations:

Not to be taken internally.
For In-Vitro Diagnostic use.
Histological applications.
Do not use if reagents become cloudy.
Do not use past expiration date.
Use caution when handling reagents.
Non-Sterile.



Control Tissue:

Any FFPE tissue.
Any Fresh or Frozen tissue.
Cell smear or spin.

Ordering Information and Current Pricing at www.scytek.com

Availability:

Item #	Volume
SAP125	125ml each
SAP500	500ml each
SAP999	1000ml each

Storage: Store at 2-8℃.

Precautions:

Avoid contact with skin and eyes.
Harmful if swallowed.
Follow all Federal, State, and local regulations regarding disposal.

Storage: 2° C 8° C

ScyTek Laboratories, Inc.
205 South 600 West
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U.S.A.

CE IVD

EC REP
Emergo Europe
Prinsessegracht 20
2514 AP The Hague, The Netherlands

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Recommended, But *Not Included*:

	Item #	Description
	PBE500	Phosphate Buffered Saline + Tween 20 (10x) pH 7.4
or	TBE500	Tris Buffered Saline + Tween 20 (10x) pH 7.5
	CPL500	Citrate Plus
	PRD500	Permanent Red Bulk Pack (For Alkaline Phosphatase)
	HMM500	Hematoxylin, Mayer's (Lillie's Modification)
	BRT500	Bluing Reagent

Procedure:

1. Rehydrate tissue slides.
2. In a glass or plastic (Autoclavable) Coplin jar, add 5 ml of Citrate Plus (CPL) and 45 ml of deionized water. *(Not included)*
3. Submerge slides in diluted Citrate Plus and loosely cap.
4. Add Distilled water to bottom of Autoclave or Pressure Cooker (about 1 inch deep in Pressure Cooker).
5. Place Coplin jar in Pressure Cooker or Autoclave.
6. Turn heat on and allow pressure to rise to 20-25 PSI.
7. Maintain pressure at 20-25 PSI for 5 minutes.
8. Turn off heat source and allow to cool.
9. When pressure has dropped to ambient, carefully remove lid or open door.
10. Using tongs, remove Coplin Jar and place on counter.
11. Once Coplin Jar cools to room temperature remove slides, rinse several times in buffer and proceed with staining as usual.
12. Apply Super Block (AAA), and incubate for 5 minutes at room temperature to block nonspecific background staining. **Note:** Do not exceed 10 minutes or there may be a reduction in desired stain.
13. Rinse 3 times in buffer.
14. Apply primary antibody and incubate according to manufacturer's protocol.
15. Rinse 3 times in buffer.
16. Apply SensiTek Anti-Polyvalent (ABF) and incubate for 20 minutes at room temperature.
17. Rinse 3 times in buffer.
18. Apply SensiTek Alk-Phos (ABM) and incubate for 20 minutes at room temperature.
19. Rinse 3 times in buffer followed by 1 rinse in DI water.

Mix Permanent Red Concentrate with Permanent Red Buffer (Not Included).

20. Add 2 drops (60ul) Permanent Red Chromogen (PRC002) to each 5ml vial of Permanent Red Buffer (PRB005), mix by swirling and apply to tissue for 5 minutes.
21. Rinse 1 time in Distilled/DI Water.
22. Apply Permanent Red Chromogen/Substrate mixture and incubate for a second 5 minute period.
23. Rinse 3 times in Distilled/DI water.
24. Apply Hematoxylin, Mayer's (HMM) and incubate for 1 minute. *(Not included)*
25 Rinse 3 times in distilled water.
26. Apply Bluing Reagent (BRT) and incubate for 5-10 seconds. *(Not included)*
27. Rinse immediately in distilled or deionized water.
28. Quickly dehydrate in alcohol and clear in xylene or substitute.

Note: Alcohol and Xylene can cause chromogen to leach from tissue over extended periods of time.

Storage: 2° C  8° C

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29. Coverslip using a permanent mounting media.

Troubleshooting Guide

Overstaining:

1. Concentration of the primary antibody was too high or the incubation time was too long.
2. Temperature during incubation was too high.
3. Incubation time with link antibody or streptavidin/enzyme label was too long.

Nonspecific Background Staining:

1. Rinsing between steps was inadequate.
2. Tissue was allowed to dry with reagents on.
3. Folds in tissue trapped reagents.
4. Tissue contains endogenous alkaline phosphatase.
5. Tissue contains endogenous biotin.
6. Antigen migrated in tissue.
7. Excessive tissue adhesive on slides.
8. Inadequate blocking with protein block.

Weak Staining:

1. Primary antibody concentration was too low or incubation time was too short.
2. Reagents are past their expiration date.
3. Inadequate removal of wash water between steps, resulting in dilution of reagents.
4. Room temperature was excessively cool.
5. The primary antibody does not recognize an antigen that survives fixation and embedding in high enough amounts.
6. Excessive incubation with protein block (Super Block).

No Staining:

1. Steps were inadvertently left out.
2. There is no antigen in the tissue.
3. The primary antibody is not of mouse, rat, rabbit or guinea pig origin.
4. Chromogenic substrate has been replaced with another that is not intended for use with alkaline phosphatase.
5. One or more components of the kit have been inactivated.

References:

1. Hsieh MH, Jan RL, Wu LS, Chen PC, Kao HF, Kuo WS, Wang JY. Lactobacillus gasseri attenuates allergic airway inflammation through PPARγ activation in dendritic cells. Journal of Molecular Medicine. 2018 Jan 1;96(1):39-51.

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