

### Instructions For Use

### UCS015-IFU

Rev. Date: Jan. 13, 2016

**Revision: 1** 

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P.O. Box 3286 - Logan, Utah 84323, U.S.A. - Tel. (800) 729-8350 - Tel. (435) 755-9848 - Fax (435) 755-0015 - www.scytek.com

# UltraTek Complete HRP Anti-Polyvalent (DAB) Staining System

**Description:** The UltraTek Complete HRP Anti-Polyvalent (DAB) Staining System provides unmatched sensitivity with

incubation times of 10 minutes each for the Link Antibody and Enzyme Label

Species of Origin: Goat

Antigen Specificity: Anti-Polyvalent (Mouse, Rat, Rabbit and Guinea Pig)

Volume

Preadsorbed Against: Human Enzyme Conjugate: Peroxidase

Chromogen Substrate: 3,3'-Diaminobenzidine (DAB)

**Uses/Limitations:** Not to be taken internally.

For Invitro Diagnostic Use. Histological applications.

Do not use if reagents become cloudy. Do not use past expiration date. Use caution when handling reagents.

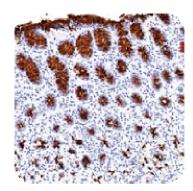
Non-Sterile.

**Control Tissue:** Any well-fixed tissue section.

Frozen tissue section. Cytocentrifuge preparation.

Ordering Information and Current Pricing at www.scytek.com

Item #



	Peroxide Block	1 x 15 ml
	Super Block	1 x 15 ml
	UltraTek Anti-Polyvalent	1 x 15 ml
	UltraTek HRP	1 x 15 ml
	DAR Chromogen	1 v 3 ml

UltraTek HRP 1 x 15 ml
DAB Chromogen 1 x 3 ml
DAB Substrate (High Contrast) 5 x 5 ml vials
Hematoxylin, Mayer's (Lillie's Mod.) 1 x 15 ml
Bluing Reagent 1 x 15 ml

**Storage:** Store at 2-8°C.

**Precautions:** Avoid contact with skin and eyes.

Harmful if swallowed.

Follow all Federal, State, and local regulations regarding disposal.

Storage: 2° C 8° C

ScyTek Laboratories, Inc. 205 South 600 West Logan, UT 84321 U.S.A.

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EC REP EmergoEurope (31)(0) 70 345-8570 Molsnstraat 15 2513 BH Hague, The Netherlands

**Kit Contents:** 



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#### Procedure:

- 1. Deparaffinize and rehydrate tissue section.
- 2. To reduce nonspecific background staining due to endogenous peroxidase, incubate slide in Peroxide Block for 10 minutes.
- 3. Wash 2 times in buffer.
- 4. If required, incubate tissue in digestive enzyme.
- 5. Wash 4 times in buffer.
- Apply Super Block (blue cap), and incubate for 5 minutes at room temperature to block nonspecific background staining.
   Note: Do not exceed 10 minutes or there may be a reduction in desired stain.
- 7. Wash 1 time in buffer.
- 8. Apply primary antibody and incubate according to manufacturer's protocol.
- 9. Wash 4 times in buffer.
- 10. Apply UltraTek Anti-Polyvalent (yellow cap), and incubate for 10 minutes at room temperature.
- 11. Wash 4 times in buffer.
- 12. Apply UltraTek HRP (red cap), and incubate for 10 minutes at room temperature.
- 13. Rinse 3 times in buffer.

WARNING: DAB is a suspected carcinogen. Handle with care and dispose of according to all regulations.

- 14. Add 5 drops (200-250ul) DAB Chromogen to one vial of DAB Substrate High Contrast, mix by swirling and apply to tissue for 5 minutes.
- 15. Rinse 1 time in distilled or deionized water.
- 16. Apply DAB Chromogen/Substrate mixture and incubate for a second 5 minute period.
- 17. Rinse 3 times in distilled or deionized Water.
- 18. Apply Hematoxylin, Mayer's (Lillie's Modification) and incubate for 1-2 minutes.
- 19. Rinse 3 times in distilled or deionized water.
- 20. Apply Bluing Reagent for 5-10 seconds.
- 21. Rinse immediately in distilled or deionized water.
- 22. Dehydrate slides and clear in xylene or xylene substitute.
- 23. Coverslip using a permanent mounting media.









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#### -Troubleshooting Guide-

#### Overstaining:

- 1. Concentration of the primary antibody was too high or the incubation time was too long.
- 2. Temperature during incubation was too high.
- 3. Incubation time with link antibody or streptavidin/enzyme label was too long.

#### Nonspecific Backgroung Staining:

- 1. Rinsing between steps was inadequate.
- 2. Tissue was allowed to dry with reagents on.
- 3. Folds in tissue trapped reagents.
- 4. Tissue contains endogenous peroxidase.
- 5. Tissue contains endogenous biotin.
- 6. Antigen migrated in tissue.
- 7. Excessive tissue adhesive on slides.
- 8. Inadequate blocking with protein block.

#### Weak Staining:

- Primary antibody concentration was too low or incubation time was too short.
- 2. Reagents are past their expiration date.
- 3. Inadequate removal of wash water between steps, resulting in dilution of reagents.
- 4. Counterstain or mounting media were incompatible and dissolved the chromogen reaction product.
- 5. Room temperature was excessively cool.
- 6. The primary antibody does not recognize an antigen that survives fixation and embedding in high enough amounts.
- 7. Excessive incubation with protein block (Super Block or normal serum).

#### No Staining:

- 1. Steps were inadvertently left out.
- 2. There is no antigen in the tissue.
- 3. The primary antibody is not of mouse, rat, rabbit or guinea pig origin.
- 4. Chromogenic substrate has been replaced with another that is not intended for use with peroxidase.
- 5. One or more components of the kit have been inactivated.

