

# Instructions For Use WGK-IFU

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## Wright-Giemsa Stain Kit

#### **Description and Principle**

Wright-Giemsa Stain Kit is intended to be used for differential staining of blood smears, bone marrow and blood parasites.

Wright-Giemsa Stain Kit uses a combination of basic and acid dyes to produce a Romanowsky-type of staining. Anionic elements are stained predominantly with the basic dyes, methylene blue and azure, whereas cationic elements are stained with the acid dye eosin.

#### **Expected Results**

Erythrocytes: Pink-Tan Leukocvtes: Blue-Purple

\*Light Purple or Lavender Neutrophils: \*Bright Red or Red-Orange Eosinophils: Basophils: \*Deep Purple or Violet-Black

\*\*Violet-Purple Platelets:

**Kit Contents Storage** 1. Wright-Giemsa Solution 18-25°C 2. Phosphate Buffer Solution (pH 6.8) 18-25°C

#### Suggested Controls (not provided)

Blood smear on clean slide.

#### **Uses/Limitations**

For In-Vitro Diagnostic use only. Do not use past expiration date. Use caution when handling reagents. Hematology Applications Non-Sterile

#### **Storage**

Store kit and all components at room temperature (18-25°C).

### Safety and Precautions

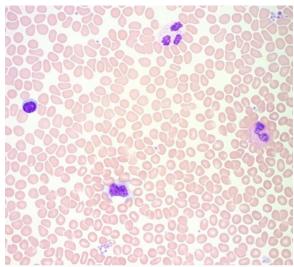
Please see current Safety Data Sheets (SDS) for this product and components GHS classification, pictograms, and full hazard/precautionary statements.

#### Preparation of Reagents Prior to Beginning:

1. Prepare Working Wright-Giemsa Solution by mixing equal parts of Wright-Giemsa Solution and Phosphate Buffer Solution, pH 6.8.

#### Procedure (Standard):

- 1. Smear a small drop of blood on a clean microscope slide and allow to air dry.
- 2. Fix by placing in absolute Methanol for 5 minutes.
- 3. Place slide in staining tray and flood with Working Wright-Giemsa Solution for 5 minutes. Note: Agitate slide occasionally to insure proper
- 4. Rinse slide in deionized/distilled water.
- 5. Flood slide with Phosphate Buffer Solution, pH 6.8 until no stain runs off.



Erythrocytes, Leukocytes, and Platelets visualized with Wright-Giemsa Stain Kit. Viewed at 400X magnification.

- 6. Allow slide to remain in Phosphate Buffer Solution, pH 6.8 for an additional 1 minute.
- 7. Dip slide in distilled water and air dry at room temperature.
- 8. Dip slide several times in Xylene or Xylene Substitute.
- 9. Mount in synthetic resin.

#### References

1. Rui Sun, Lixiazi He, Hyeyoon Lee, Andrey Glinka, Carolin Andresen, Daniel Hübschmann, Irmela Jeremias, Karin Müller-Decker, Caroline Pabst, Christof Niehrs, RSPO2 inhibits BMP signaling to promote self-renewal in acute myeloid leukemia, Cell Reports, Volume 36, Issue 7, 2021, 109559, ISSN 2211-1247, https://doi.org/10.1016/j.celrep.2021.109559.

2. Wang, J., Li, R., Peng, Z., Hu, B., Rao, X., & Li, J. (2020). HMGB1 participates in LPS-induced acute lung injury by activating the AIM2 inflammasome in macrophages and inducing polarization of M1 macrophages via TLR2, TLR4, and RAGE/NF-kB signaling pathways Corrigendum in /10.3892/ijmm.2020.4530. International Journal of Molecular Medicine, 45, 61-80. https://doi.org/10.3892/ijmm.2019.4402

3. Sheehan, D., Hrapchak, B., Theory and Practice of Histotechnology: 2nd Edition, 1980, pages 155-156.

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IVD

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<sup>\*</sup>granules in cytoplasm.

<sup>\*\*</sup>granules in light blue cytoplasm